

# Some hemato-biochemical, bacteriological and pathological effects of *Moringa oleifera* leaf extract in broiler chickens

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## Abstract

Cloacal swabs were collected from 300 broiler chicks (50 dead, 50 diseased and 200 healthy) for bacteriological examination. Out of 300 examined cloacal swabs 26 (52%) dead, 28 (56%) diseased and 28 (14%) from healthy chicks were positive for bacteria. *Moringa oleifera* leaf extract inhibits growth of *E. coli* and *Salmonella* spp. Post bacteriological examination 120 chicks free from bacterial infection, one day old were divided into 3 equal groups. 1st group healthy chicks (control), 2nd and 3rd groups chicks received 200 mg/ kgm bwt watery and alcoholic moringa oleifera leaf extract daily for 30 successive days in drinking water respectively. Effect of moringa leaf extract on body weight, feed conversion rate and haemato-biochemical analysis beside pathological changes of some internal organ were study.

Moringa leaf extracts (watery & alcoholic) induced significant increase in body weight gain, RBCs, Hb, PCV% WBCs, total proteins, albumin, globulins, SOD beside significant decrease in MDA and insignificant increase in AST, ALT, ALP but urea and creatinine insignificantly decreased and improved in feed conversion rate.

Histopathological investigation revealed, Hepatic tissue, congestion and hyperplasia in bile duct epithelium. Renal tubules undergo interstitial aggregation of lymphocytes and regeneration. Intestine shows slight edema. spleen and bursa show hyperplasia of lymphocyte in white pulp.

It could be concluded that Moringa leaf extract act antibacterial, growth promoter, antioxidant and have benefits effect in immunity and hemato-biochemical parameters.

**Keywords:** *Moringa Oleifera*; Leaf Extract; Broiler Chicken; Histopathology.

## 1. Introduction

Chicken industry is one of the most dynamic of world agribusiness trade. Hence, research on meat production globally indicates poultry as fastest growing livestock sector especially in developing countries over several years; antibiotics are widely used in chicken for therapeutic use to treat diseases, prophylactic use to prevent infections or as growth promoters. However, problems associated with the use of antibiotics in poultry production include drug toxicity and residual effect [1]. Extensive use of antibiotics renders most antimicrobial agents inefficient in controlling some bacterial diseases [2].

*Moringa oleifera* plant possesses medicinal properties [3]. Moringa oleifera leaf extract has antibacterial properties [4]. Moringa oleifera was claimed to boost immune systems [5]. Its exhibit antitumor, anti-inflammatory and antiulcer activities [6]. Moringa oleifera are rich sources for natural anti-oxidants as tocopherols, flavonoids and vitamin C and essential oils [7]. Moringa oleifera have several advantages over commonly used commercial antibiotics, since they are residue free and are generally recognized as safe and commonly used in the food industry [8]. The aim of the present study is to investigate the antibacterial ability of Moringa oleifera leaf extract against some pathogenic bacteria, its effect in body performance, some hematobiochemical

parameters, some antioxidant enzymes as well as pathological changes in some internal organ in broiler chickens

## 2. Material and methods

### 2.1. Bacteriological examination

Cloacal swabs were collected from 300 chicks, one day old (50 dead, 50 diseased and 200 healthy). All swabs were collected aseptically, inoculated into nutrient broth aerobically at 37°C overnight, subculturing on nutrient agar and MacConkey agar plates, incubated for 24h at 37°C, colonies were identified [9].

### 2.2. Serological identification

The isolated *E. coli* and *Salmonella* species strains were serotyped using slide agglutination test against polyvalent and monovalent standard serum obtained from Denka sickon company LTD Tokyo Japan, using methods of [10].

Collection and Preparation of Aqueous and Ethanolic Extract  
Leaf of moringa oleifera was collected; air dried and pounded using mortar and pestle prior to extraction (watery and ethyl alcohol) [11].

### 2.3. Antibiotic sensitivity test (in vitro)

Susceptibility of isolated Salmonella spp and E. coli to moringa oleifera leaf extract in comparison to some chemotherapeutic agents was tested by disc diffusion method [12].

### 2.4. Birds and experimental design

Post bacteriological examination for 200 healthy chickens nearly equal in the live body weight (38-40gm), 120 chicks free from any bacterial infection were divided into 3 groups (40 chicks each), 1<sup>st</sup> group served as control group. 2<sup>nd</sup> and 3<sup>rd</sup> groups supplemented with 200 mg/kgm bwt watery and alcoholic extract of moringa oleifera leaf in drinking water respectively for 30 days (from 1<sup>st</sup> day of age up to 30<sup>th</sup> day of age)

### 2.5. Body weight

From each group 5 chicks were weighted individually at the start of the experiment and at 1<sup>st</sup> day post supplementation (corresponding 31<sup>th</sup> day of age) and consumed diets were recorded for calculation of weight gain and feed conversion rate.

### 2.6. Blood samples

At 1<sup>st</sup> and 10<sup>th</sup> day post supplementation 5 chicks from each group were sacrificed and two blood samples were collected. First sample was taken in tube containing EDTA as anticoagulant for hematological studies according [13], superoxide dismutase (SOD) determined spectrophotometrically [14]. Malondialdehyde (MDA) [15], 2<sup>nd</sup> sample was used for obtaining clear serum for estimation

total protein [16], albumin [17], globulin was calculated as difference between total protein and albumin, (AST & ALT) [18], ALP [19], uric acid [20] creatinine [21].

### 2.7. Pathological examination

Specimens were taken from liver, kidneys, intestine, spleen and bursa of the sacrificed chicks and directly fixed in 10% neutral buffered formalin. Five micron thick paraffin sections were prepared, stained with hematoxylin and eosin and examined microscopically [22]. Statistical analysis of the obtained data was analyzed (T test) [23].

## 3. Results

Bacteriological and hematobiochemical results were recorded in tables (1-8)

*Moringa oleifera* leaf extract addition : Liver shows hyperplasia in bile duct epithelium of watery extract (Fig 1), alcoholic extract shows liver congestion (Fig 2), kidney of watery extract shows interstitial aggregation of lymphocytes (Fig. 3) an alcoholic extract shows regenerative attempts in the renal tubules (Fig 4), Intestine of alcoholic extract shows slight edema (Fig 5) Bursa of watery extract shows hyperplasia in constituent cells (Fig. 6) and alcoholic extract shows severe hyperplasia in bursal lymphocytes (Fig.7), Spleen of watery moringa extracts showing thickening of splenic arterioles (arrow) and depletion of white pulp (arrowhead) (Fig 8), Spleen of alcoholic-moringa extract shows hyperplasia in the lymphocytes of white pulp (Fig 9).

**Table 1:** Prevalence of Bacterial Isolates from Cloacae Swabs.

Swabs	No. of swabs	Single isolates		Mixed isolates		Total +ve		Total -ve	
		No	%	No	%	No	%	No	%
dead	50	12	24	14	28	26	52	24	48
diseased	50	15	30	13	26	28	56	22	44
healthy	200	11	5.5	17	8.5	28	14	172	86

**Table 2:** Incidence of Different Bacterial Pathogens Isolated from Examined Cloacal Swabs.

Swabs origin	Swabs number	Isolates		Isolated organisms	No	%	
		Type	No				
Dead chicks	50	Single	12	24	E.coli	5	41.66
					Proteus	3	25
					Salmonella spp.	2	16.66
					Staph. Aureus	2	16.66
		Mixed	14	28	Pseudomonas+ E.coli+ Proteus	5	35.71
					Staph. Aureus+ klebsiella	8	57.14
Diseased chicks	50	Single	15	30	E.coli	6	40
					Salmonella spp.	1	6.67
					Strept. spp.	4	26.67
		Mixed	13	26	Proteus + klebsiella	5	38.46
					Staph. Aureus+ E.coli	3	23.08
					Staph. Aureus+ Proteus	5	38.46
Healthy chicks	200	Single	11	5.5	E.coli	4	36.36
					Salmonella spp.	2	18.18
					Pseudomonas	3	27.27
		Mixed	17	8.5	E coli + Staph. Aureus	6	35.29
					Proteus + Staph. Aureus+ E coli	5	29.41
					Salmonella spp.+ Proteus	4	23.52
Salmonella spp + E.coli	2	11.76					

**Table 3:** Serological Identification of Isolated E.Coli and Salmonella Strains.

Isolated bacteria	serotyp	Dead (5)	Diseased (6)	Healthy (4)	Total
E.coli	O157	1	2	1	4
	O78	2	1	2	5
	O11	1	3	1	6
Salmonella		Dead (2)	Diseased (1)	Healthy (2)	
	S. entritidis	-	1	1	2
	S. typhimurium	2	-	1	3

**Table 4:** In-Vitro Antibacterial Effect of Moringa Olefera on Some Isolated Bacteria.

Bacteria Parameter		E.coli		Salmonella spp	
		Inhibition zone(mm)	Sensitive	Inhibition zone(mm)	Sensitive
Moringa Olefera	watery	10.17	++	13.14	++
	alcoholic	11.38	++	13.95	++
Florphenicol (FF)		20	+++	19.83	+++
Gentamycin (GM 10)		15	++	13.39	++
Kanamycin (K 30)		14	++	14.32	++

**Table 5:** Effect of Moringa Olefera in Body Weight of Broiler (N=5).

Period&groips Parameter	1 <sup>st</sup> day			10 <sup>th</sup> day		
	control	Watery	alcoholic	control	Watery	alcoholic
Initial body weight	39.83 ±0.27	40.01 ±0.38	38.97 ±0.22	39.88 ±0.24	39.07 ±0.25	40.05 ±0.24
Final body weight	1420.75 ±5.34	1530.32 ±7.84	1557.40 ±6.38	1974.69 ±5.83	2005.42 ±6.95	2015.55 ±8.43
Weight gain	1380.92 ±6.42	1490.25 ±4.81	1518.43 ±5.48	1934.81 ±6.53	1966.35 ±5.49	1975.50 ±8.43
Feed consumption	1640.42 ±2.97	1696.06 ±3.44	1503.18 ±3.81	2603.07 ±2.57	2615.53 ±4.83	2010.04 ±3.72
FCR	1.19	1.12	0.99	1.35	1.33	1.02

FCR= Feed Conversion rate \* Significant at P &lt; 0.05 \*\* Significant at P &lt; 0.001

**Table 6:** Effect of Moringa Olefera in Blood Picture, Phagocytosis & Killing % in Broiler (N=5).

Period&groips Parameter	1st day			10 <sup>th</sup> day		
	control	watery	alcoholic	control	Watery	alcoholic
RBCs (10 <sup>6</sup> /mm <sup>3</sup> )	3.35± 0.17	3.95± 0.14*	4.04± 0.21*	3.41± 0.21	3.58± 0.19	3.79± 0.26
HB (gm/dl)	13.24± 0.20	14.23± 0.34*	14.74± 0.52*	13.25± 0.32	13.53± 0.29	14.12± 0.40
PCV%	34.28± 0.86	37.22± 0.94*	37.85± 0.89*	34.60± 0.94	35.17± 0.78	34.69± 0.98
WBCs (U/L)	11.31± 0.74	13.13± 0.16*	13.34± 0.22*	11.52± 0.89	12.49± 0.31	12.08± 0.52
Phagocytosis	54.09± 1.21	58.16± 1.37*	58.62± 1.42*	54.16± 1.27	57.03± 1.23	58.06± 1.55
Killing %	43.19± 1.22	44.84± 1.05	45.04± 1.13	43.31± 1.26	44.15± 1.32	44.21± 1.40

\* Significant at P &lt; 0.05

**Table 7:** Effect of Moringa Olefera on Some Biochemical Parameters in Broiler (Mean ± S.E.)

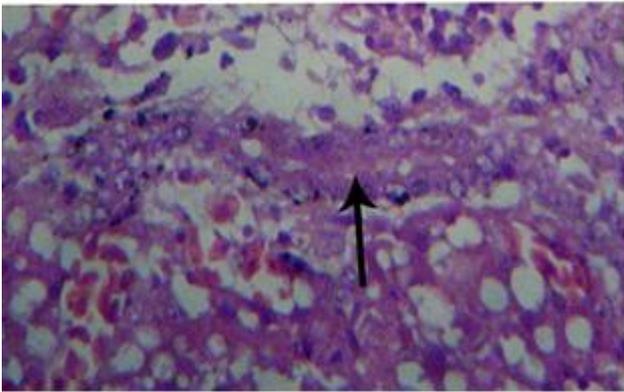
Period& groips Parameter		1 <sup>st</sup> day			10 <sup>th</sup> day		
		control	watery	alcoholic	Control	watery	alcoholic
Protein picture (mg/dl)	T.Protein	5.12± 0.32	6.44± 0.38*	6.63± 0.43*	5.19± 0.28	5.68± 0.41	5.92± 0.39
	Albumin	2.99± 0.20	3.65± 0.19*	3.77± 0.20*	2.90± 0.24	3.21± 0.22	3.20± 0.35
	Globulin	2.13± 0.21	2.79± 0.17*	2.86± 0.14*	2.29± 0.23	2.47± 0.19	2.72± 0.21
	A/G	1.40± 0.18	1.31± 0.13	1.32± 0.21	1.27± 0.18	1.30± 0.17	1.32± 0.15
Liver Enzymes )U/L(	AST	49.38± 1.05	49.67± 1.48	49.70± 1.93	49.50± 1.24	49.62± 1.61	59.72± 1.50
	ALT	63.71± 1.48	64.03± 1.61	64.08± 1.52	63.47± 1.59	63.53± 1.36	63.49± 1.68
	ALP	43.21± 1.85	43.45± 1.25	43.52± 1.69	43.18± 1.93	43.24± 1.21	43.28± 1.88
Kidney Function )mg/dl(	Uric acid	5.36± 0.41	5.19± 0.43	5.27± 0.31	5.30± 0.30	4.21± 0.26	4.24± 0.52
	Creatinine	1.15± 0.10	1.12± 0.13	1.10± 0.18	1.17± 0.15	1.14± 0.27	1.13± 0.12

\* Significant at P &lt; 0.05

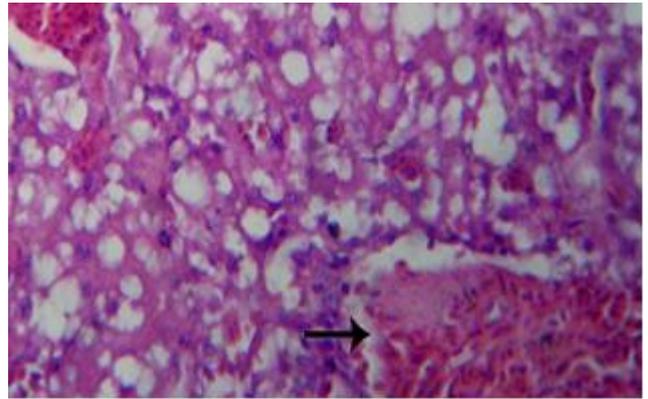
**Table 8:** Effect of Moringa Olefera on Some Antioxidant Enzymes in Broiler (Mean ± S.E.)

Period& groips Parameter	1st day			10th day		
	control	Watery	alcoholic	Control	watery	alcoholic
SOD (U/ml)	236.14± 2.59	244.09± 1.85*	245.13± 1.95*	238.95± 2.61	242.16± 1.69	243.25± 1.95
MDA (nmol/ml)	14.38± 0.89	11.15± 0.97*	10.89± 0.88*	14.64± 0.68	13.32± 0.88	13.06± 0.96

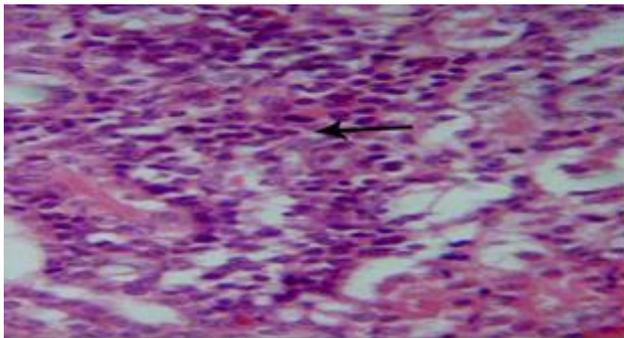
\* Significant at P &lt; 0.05



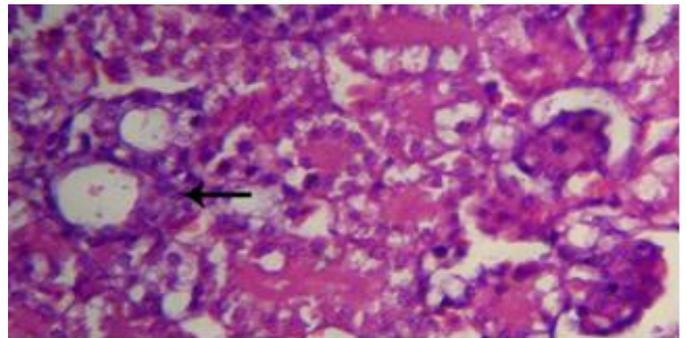
**Fig 1:** Liver of Broiler Chicken in Watery -Moringa Leaf Extract Showing Hyperplasia in Bile Duct Epithelium (HE X 400)



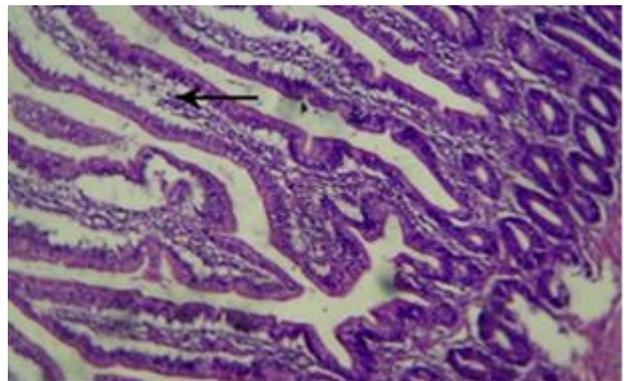
**Fig 2:** Liver of Broiler Chicken in Alcoholic-Moringa Extract Showing Congestion(HeX400)



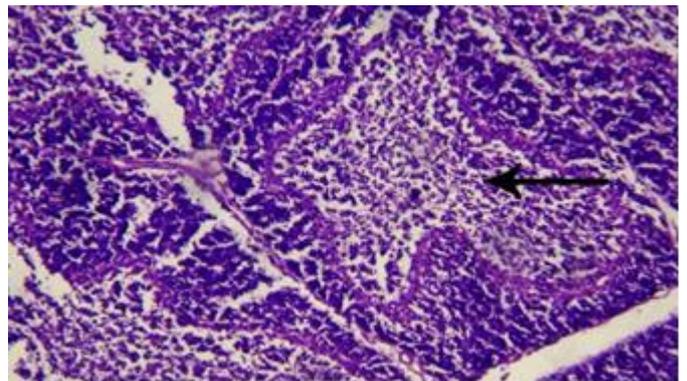
**Fig 3:** Kidney of Broiler Chicken in Watery -Moringa Extract Showing Interstitial Aggregation of Lymphocytes. (He X 400)



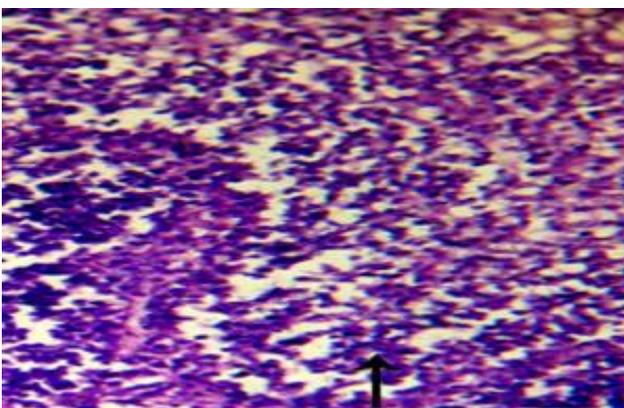
**Fig 4:** Kidney of Broiler Chicken in Alcoholic-Moringa Extracts Showing Regenerative Attempts in the Renal Tubules (He X 400)



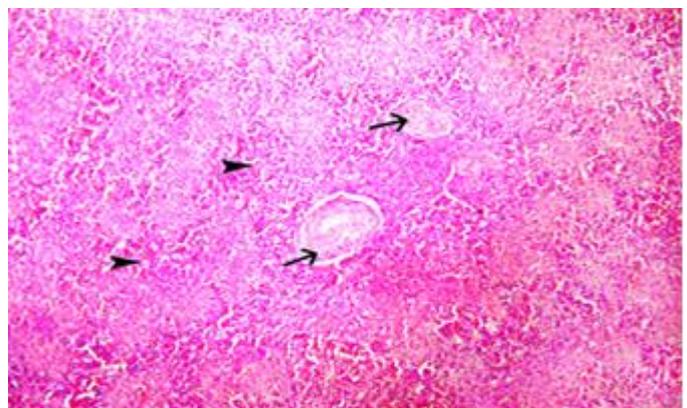
**Fig 5:** Intestine of Broiler Chicken in Alcoholic-Moringa Extract Showing Slight Edema. (HE X 200).



**Fig 6:** Bursa of Broiler Chicken in Watery Moringa Olefera Extract Showing Hyperplasia in the Constituent Cells.(He X 200)



**Fig 7:** Bursa of Broiler Chicken in Alcoholic-Moringa Extract Showing Severe Hyperplasia in the Bursal Lymphocytes. (HE X 400).



**Fig 8:** Spleen of Broiler Chicken in Watery Moringa Olefera Extracts Showing Thickening of Splenic Arterioles (Arrow) and Depletion of White Pulp (Arrowhead), H E X300.

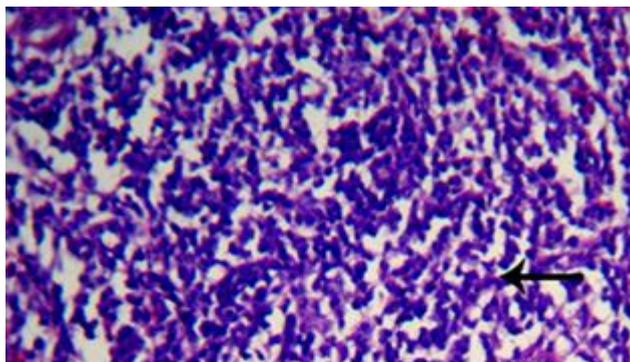


Fig 9: Spleen of Alcoholic-Moringa Extract Shows Hyperplasia in the Lymphocytes of White Pulp. (HE X 400).

#### 4. Discussion

Out of 300 examined swabs 26 (52%) dead, 28 (56%) diseased and 28 (14%) from healthy chicks were positive for bacteria either single or mixed isolants (table 2). Our data are in accordance with [24] who isolates *E. coli*, salmonella spp., *Pseudomonas*, *Proteus* and these bacteria may be found in single or mixed infections. *Salmonella* spp. Were isolated from diseased broiler chickens [25]. Our results agreed with results recorded by [26] isolate *Salmonella* species and *E. coli* from initial few weeks of chicks life.

The isolated *E. coli* (15) were differentiated into 3 typed isolates {O11 (6), O78 (5) and O157 (4)} serotypes (table 3). similar results recorded [27]. Serological identification of *Salmonella* isolates revealed that *Salmonella* enteritidis (2) and *Salmonella* typhimurium (3) was the most serotype in our study. same results were obtained by [28] isolate *Salmonella* enteritidis, *Salmonella* Gallinarum and *Salmonella* typhimurium were the commonly isolated serotypes from broiler flocks.

In the present study, *Moringa oleifera* leaf extracts inhibits growth of *Salmonella* species and *E. coli*. alcoholic leaf extract induced zone of inhibition more than watery extract (table, 4). This finding was supported by [4] stated that *E. coli* sensitive to *Moringa oleifera* leaf extract. Ethanoic leaf extract of *Moringa oleifera* induced zone of inhibition against *Salmonella typhii* more than inhabitation zone of *E. coli* [29]. *Moringa oleifera* extract have antibacterial properties [30]. *Moringa oleifera* leaf extract used as alternatives to antibiotics [7]. Medicinal plants as *Moringa oleifera* may represent an alternative treatment for non severe cases of infectious diseases [31].

Healthy broiler chicks received *Moringa* leaf extract (watery and alcoholic) resulted a significant increase in body weight, weight gain and improved in feed conversion rate (table, 5). These obtained results go in agreement with [32], [33] and [34] in broiler chickens supplemented with moringa extract. varying concentrations of moringa aqueous leaf extract induce increase in body weight gain and improved in feed conversion rate in Cobb broilers [35]. Elevation in weight gain and improved in feed conversion rate may be attributed to presence of high amounts of vitamins, minerals and amino acids in *Moringa* leaf [36].

In the current work, broiler chickens supplemented with 200 mg/kgm bwt *Moringa* leaf extract (watery and alcoholic) induce significant increase in total erythrocytic count, haemoglobin content, packed cell volume, total leukocytic count, phagocytosis and killing% at 1<sup>st</sup> day post administration (table, 6). Same result was reported by [37] in broiler chickens received *Moringa* leaf extract. Elevation in erythrocytic count, haemoglobin content and packed cell volume % post using *Moringa* may be due to presence of saponin in *Moringa* [38] and/or alkaloids and flavonoids in *Moringa* induce increase in total erythrocytic count, haemoglobin content and packed cell volume % [39]. *Moringa* in dose of 100-400 mg/kg induced increase in leukocytic counts [40]. *Moringa oleifera* induce improvement in phagocytosis and killing % [41 and 42] in broilers chickens and rats respectively.

The obtained results revealed significant increase in serum total proteins, albumin and globulins at 1<sup>st</sup> day post supplementation of 200 mg /kgm bwt. *Moringa* leaf extract in drinking water to broil-

er chickens (table, 7). Our results were coinciding with those obtained by [39] who stated that moringa induce significant increase in total protein and albumin. Same result was recorded by [1-34] in broiler chickens received moringa. Elevations in total protein our study may be due to moringa are rich in both essential and sulfur-containing amino acids [43] and/ or due to *Moringa oleifera* contain large amount of protein [44].

In the present experiment, *Moringa* leaf extract in a dose of 200 mg / kgm bwt induces insignificant increase in AST, ALT and ALP beside insignificant decrease in urea and creatinine (table, 7). Same changes were observed [5] in broilers fed *Moringa* leaf. *Moringa* leaf extract induce insignificant effect in liver enzymes, urea and creatinine [45]. Also, [41] reported *Moringa* extract induce non significant decreases in uric acids of broilers. This obtained result was similar to those recorded [46] who found that *Moringa* leaf extract induce non significant effect in liver enzymes. *Moringa* leaf extract have hepatoprotective effect [47].

In this study, the superoxide dismutase showed significant increased beside significant decrease in malondialdehyde in broiler chickens received *Moringa* leaf extract (table, 8). Same changes were recorded [48] in laying hens received *Moringa* leaf extract. *Moringa* contain antioxidant enzymes, reducing intensity of lipid peroxidation and inhibiting generation of free radicals [49-50]. The antioxidant effect of *Moringa* leaf extract may be due to presence of polyphenols, anthocyanin, thiocarbamates and glycosides which remove free radicals, activate antioxidant enzymes and inhibit oxidases [51] and/ or due to *Moringa* suppress formation of reactive oxygen species and free radicals [52]

The histomorphological changes in Liver was hyperplasia in bile duct epithelium and congestion post using moringa extract. Same microscopical lesions were recorded by [53]. kidney showed interstitial aggregation of lymphocytes and regenerative attempts in the renal tubules. These findings were supported by [46]. Intestine showed slight edema as recorded by [45]. In the immune organs (bursa and spleen) showed hyperplasia of lymphocytes. The microscopic lesions recorded in the present investigation were almost similar to the lesions recorded by [37].

It could be concluded that *Moringa oleifera* leaf extract act as antibacterial, growth promoters, antioxidant and exhibits some benefits effect on immunity and hemato-biochemical parameters.

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